## STRUCTURAL MODIFICATION OF LIGNAN COMPOUNDS ISOLATED FROM NECTANDRA SPECIES (LAURACEAE)

# VÍCTOR MACÍAS-VILLAMIZAR<sup>1,1\*,§</sup>, LUÍS CUCA-SUÁREZ<sup>1</sup>

<sup>1</sup>Departamento de Química, Laboratorio de Productos Naturales, Universidad Nacional de Colombia. Av. Carrera 30 # 45-03. Cód. postal 111321; Edificio 476-Oficina 11; Bogotá D.C., Colombia. §Profesor Universidad del Magdalena, Colombia.

### **1-INTRODUCTION**

In the genus *Nectandra*, the presence of certain types of secondary metabolites has been determined, including sesquiterpenes, phytosterols, polyalcohols, arylpropionic acid derivatives, flavonols, arylpropanoids, furofuran lignans, dihydrobenzofuran neolignans [1], and certain norlignans [2], alkaloids [3], tannins [4], diterpenes [5], and components of essential oils [6]. However, the chemotaxonomic characteristics are determined by the presence of lignan-type compounds [7]. The ultimate goal of structural modification of natural products is to obtain new drugs [8]. In that sense, there is a growing interest in lignans and their synthetic derivatives due to applications in cancer chemotherapy and various other pharmacological effects [9]. This work corresponds to the first report of this type of structural modification of lignan compounds (7,7'-epoxylignans and diaryldimethylbutane lignans) isolated from *Nectandra* species. Therefore, this work can be used as a starting point for structure-activity relationship studies.

#### **EXPERIMENTAL**

#### Materials and reagents

Benzyl bromide (Merck), acetone and toluene were freshly distilled before use. 2,3-Dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) (Fluka purum ,97%), 1,4-dioxane (Aldrich, anhydrous 99.8%), pyridine (ACS reagent,  $\geq$ 99.0%, Sigma-Aldrich) and acetic anhydride (ACS reagent,  $\geq$ 98.0%, Sigma-Aldrich) were used. Purification of the products was carried out on a short silica gel column (100–200 mesh, Merck) using increasing percentage of ethyl acetate in hexane as elutant. NMR spectra: were recorded on a Bruker Avance 400 spectrometer (<sup>1</sup>H 400 MHz, <sup>13</sup>C 100 MHz) using TMS as internal standard, in deuterated chloroform (CDCl<sub>3</sub>) as solvent. The product ethers were characterized by comparing spectral data of known compounds described in the literature and analysis of the spectral data.



Figure 1. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) Spectrum of Compound 1 (mixture of veraguensin/galgravin)

*Veraguensin*: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$ : 0.67 (3H, *d*, *J* = 7.0, H-9<sup>3</sup>), 1.07 (3H, *d*, *J* = 6.6, H-9), 1.79 (1H, *m*, H-8), 2.25 (1H, *m*, H-8<sup>3</sup>), 3.86 (*s*, OCH<sub>3</sub>), 3.88 (*s*, OCH<sub>3</sub>), 3.89 (*s*, OCH<sub>3</sub>), 3.91 (*s*, OCH<sub>3</sub>), 4.42 (1H, *d*, *J* = 9.3, H-7), 5.14 (1H, *d*, *J* = 8.6, H-7<sup>3</sup>), 6.86-7.08 (6H, *m*, H-2/5/6, H-2<sup>3</sup>/5<sup>3</sup>/6<sup>3</sup>). [**v**=veraguensin].

*Galgravin:* <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$ : 1.05 (6H, d, J = 6.7, H-9/9'), 2.34 (2H, m, H-8/8'), 3.87 (s, OCH<sub>3</sub>), 3.88 (s, OCH<sub>3</sub>), 4.52 (2H, d, J = 6.4, H-7/7'), 6.85-6.99 (6H, m, H-2/5/6, H-2'/5'/6'). [g=galgravin].



Figure 2. <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) Spectrum of Compound 1 (mixture of veraguensin/galgravin)

*Veraguensin:* <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 149.1 (C), 148.7 (C), 148.7 (C), 148.2 (C), 133.9 (C), 133.6 (C), 119.3 (CH), 118.8 (CH), 111.1 (CH), 110.8 (CH), 110.5 (CH), 110.1 (CH), 87.4 (CH), 83.1 (CH), 56.1 (CH<sub>3</sub>), 56.0 (CH<sub>4</sub>) (x2), 55.9 (CH<sub>4</sub>), 48.0 (CH), 46.1 (CH), 15.2 (CH<sub>4</sub>), 15.1 (CH<sub>3</sub>).

*Galgravin:* <sup>13</sup>C NMR (100 MHz) δ: 12.9 (CH<sub>3</sub>), 44.3 (CHCH<sub>3</sub>), 55.8 (OCH<sub>3</sub>), 55.9 (OCH<sub>3</sub>), 87.2 (OCH(Ar)), 109.7 (Ar-C2), 110.9 (Ar-C5), 118.5 (Ar-C6), 134.8 (Ar-C1), 148.4 (Ar-C4), 148.9 (Ar-C3).

#### **Extraction of Secondary Metabolites**

Secondary metabolites were previously isolated from species of *Nectandra sp.* in the Natural Products Laboratory of the National University of Colombia and correspond to 7,7'-epoxilignan and diaryldimethylbutane lignans.

#### **General Procedure**

#### Aromatization of veraguensina and galgravin (1).

A solution of veraguensin and galgravin (90/10) **1**, (5,0 mmol) and DDQ (15,0 mmol) in toluene (20 mL) (the reaction mixture immediately turned deep green) was refluxed  $(100^{\circ}\text{C})$  for 6 h. The mixture was cooled, the precipitate collected, the solvent evaporated under reduced pressure, and the resulting residue purified by flash chromatography on silica gel (n-hexane/AcOEt= 7/3) to give 2,5-bis(3,4-dimethoxyphenyl)-3,4-dimethylfuran (45%) (**TL-1**) [10].

<sup>1</sup>*H* NMR (400 MHz, CDCl<sub>2</sub>) spectral data of Veraguensin: δ: 0.67 (3H, d, J = 7.0, H-9'), 1.07 (3H, d, J = 6.6, H-9), 1.79 (1H, m, H-8), 2.25 (1H, m, H-8'), 3.86 (s, OCH<sub>3</sub>), 3.88 (s, OCH<sub>3</sub>), 3.89 (s, OCH<sub>3</sub>), 3.91 (s, OCH<sub>3</sub>), 4.42 (1H, d, J = 9.3, H-7), 5.14 (1H, d, J = 8.6, H-7'), 6.86-7.08 (6H, m, H-2/5/6, H-2'/5'/6').

<sup>1</sup>*H* NMR (400 MHz, CDCl<sub>3</sub>) spectral data of Galgravin: δ: 1.05 (6H, d, J = 6.7, H-9/9'), 2.34 (2H, m, H-8/8'), 3.87 (s, OCH<sub>3</sub>), 3.88 (s, OCH<sub>3</sub>), 4.52 (2H, d, J = 6.4, H-7/7'), 6.85-6.99 (6H, m, H-2/5/6, H-2'/5'/6').

<sup>1</sup>*H* NMR (400 MHz, CDCl<sub>3</sub>) spectral data of **TL-1**:  $\delta$  7.21 (4H, dd, J = 6.1, 1.9, H-2/2' and H-6/6'), 6.94 (2H, d, J = 8.9, H-5/5'), 3.95 (3H, s, OCH<sub>3</sub>-3/3'), 3.92 (3H, s, OCH<sub>3</sub>-4/4'), 2.22 (s, 6H, H-9/9') (see supporting information, Figure 3).



**Figure 3.** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) Spectrum of Compound 1 modified (**TL-1**) (mixture of veraguensin/galgravin modified)

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.21 (4H, *dd*, *J* = 6.1, 1.9, H-2/2' and H-6/6'), 6.94 (2H, *d*, *J* = 8.9, H-5/5'), 3.95 (6H, *s*, OCH<sub>3</sub>-3/3'), 3.92 (6H, *s*, OCH<sub>3</sub>-4/4'), 2.22 (6H, *s*, H-9/9').



**Figure 4.** <sup>13</sup>C NMR (400 MHz, CDCl<sub>3</sub>) Spectrum of Compound 1 modified (**TL-1**) (mixture of veraguensin/galgravin modified)

<sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 149.2 (C) (x2), 148.3 (C) (x2), 147.1 (C) (x2), 125.3 (C) (x2), 118.6 (CH) (x2), 117.9 (C) (x2), 111.5 (CH) (x2), 109.5 (CH) (x2), 56.2 (CH<sub>3</sub>), 56.1 (CH<sub>3</sub>), 10.0 (CH<sub>3</sub>) (x2).



Figure 5. COSY Spectrum of Compound 1 modified (TL-1) (mixture of veraguensin/galgravin modified)



**Figure 6.** Expansion of the COSY Spectrum of Compound 1 modified (**TL-1**) (From 4.80 to 3.40  $_{\rm f1}$  and From 7.45 to 6.65  $_{\rm f2}$ ) (mixture of veraguensin/galgravin modified)



**Figure 7.** Expansion of the COSY Spectrum of Compound 1 modified (**TL-1**) (From 7.60 to 6.70  $_{\rm f1}$  and From 7.36 to 6.88  $_{\rm f2}$ ) (mixture of veraguensin/galgravin modified)



Figure 8. HMQC Spectrum of Compound 1 modified (TL-1) (mixture of veraguensin/galgravin modified)



**Figure 9.** Expansion of the HMQC Spectrum of Compound 1 modified (**TL-1**) (From 124.0 to 102.0  $_{\rm fl}$  and From 7.34 to 6.90  $_{\rm f2}$ ) (mixture of veraguensin/galgravin modified)



**Figure 10.** Expansion of the HMQC Spectrum of Compound 1 modified (**TL-1**) (From 70.0 to 0.0  $_{r1}$  and From 4.20 to 1.80  $_{r2}$ ) (mixture of veraguensin/galgravin modified)



Figure 11. DEPT-135° Spectrum of Compound 1 modified (TL-1) (mixture of veraguensin/galgravin modified)



Figure 12. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) Spectrum of Compound 2 (schineolignin B)

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.76 (2H, *d*, *J* = 8.1, ArH), 6.65 (1H, *d*, *J* = 1.9, ArH), 6.63 (1H, *dd*, *J* = 8.1, 1.8, ArH), 6.58 (1H, *d*, *J* = 1.8, ArH), 2.56 (2H, *dd*, *J* = 13.5, 6.8, H-7/7<sup>°</sup>), 2.40 (2H, *dd*, *J* = 13.7, 7.8, 7/7<sup>°</sup>), 1.76 (2H, *dd*, *J* = 12.9, 6.5, H-8/8<sup>°</sup>), 0.83 (6H, *d*, *J* = 6.6, H-9/9<sup>°</sup>).

#### Acetylation of schineolignin B (2).

Schineolignin B 2 (2,1 mmol), in a mixture of acetic anhydride and pyridine (5 mL/ 5 mL) was placed in a 50 mL pear-shaped flask. The mixture was stirred at 100 C for 15 h. Removal of the solvent under reduced pressure afforded a crude mixture, which was extracted with HCl solution followed by extracted with NaHCO<sub>3</sub> solution to give the products, which was purified by column chromatography on silica gel (n-hexane/AcOEt= 8/2) *Sephadex LH-20* in open column chromatography to give 5-(4-(3,4-dimethoxyphenyl)-2,3-dimethoxyphenyl acetate, 85% (**TL-2**) [11].

<sup>1</sup>*H* NMR (400 MHz, CDCl<sub>3</sub>) spectral data of Schineolignin B: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 6.76 (1H, *d*, *J* = 8.1, Ar-H), 6.65 (1H, *d*, *J* = 1.9, Ar-H), 6.63 (1H, *dd*, *J* = 8.1, 1.8, Ar-H), 6.58 (1H, *d*, *J* = 1.8, Ar-H), 2.56 (2H, *dd*, *J* = 13.5, 6.8, H-7/7<sup>2</sup>), 2.40 (2H, *dd*, *J* = 13.7, 7.8, H-7/7<sup>2</sup>), 1.76 (2H, *dd*, *J* = 12.9, 6.5, H-8/8<sup>2</sup>), 0.83 (6H, *d*, *J* = 6.6, H-9/9<sup>2</sup>).

<sup>1</sup>*H* NMR (400 MHz, CDCl<sub>3</sub>) spectral data of **TL-2**: δ: 6.76– 6.57 (5H, m, Ar-H), 3.86–3.81 (12H, s, 3 x OCH<sub>3</sub>), 2.56 (2H, dd, *J* = 13.5, 6.7, H-7/7<sup>'</sup>), 2.40 (2H, dd, *J* = 13.5, 7.8, H-7/7<sup>'</sup>), 2.30 (3H, s, CH<sub>3</sub>-CO<sub>2</sub>-Ar), 1.79–1.73 (m, 2H, H-8/8<sup>'</sup>), 0.83 (d, *J* = 6.6, 6H, H-9/9<sup>'</sup>) (see supporting information, Figure 13).

# Benzylation of *meso*-dihydroguaiaretic acid and *threo*-dihydroguaiaretic acid (3).

A mixture of **3**(0.567 mmol) and sodium carbonate (11.4 mmol) in dry acetone (36 ml) was heated to reflux for 1 h under nitrogen. Then, benzyl bromide (0.63 ml, 5.67 mmol) was added and the mixture was heated under reflux for an additional 3 h. After cooling to room temperature, the reaction mixture was filtered. The filtrate was concentrated and distilled under reduced pressure in a rotary evaporator to remove the excess unreacted benzyl bromide. The residue was chromatographed on silica gel (hexane/AcOEt= 8/2) and *Sephadex LH-20* in open column chromatography to give 1-(benzyloxy)-4-(4-(3,4-dimethoxyphenyl)-2,3-dimethylbutyl)-2-methoxybenzene, 80% (TL-3) [12, 13].

<sup>1</sup>*H* NMR (400 MHz, CDCl<sub>3</sub>) spectral data of meso-dihydroguaiaretic acid and threo-dihydroguaiaretic acid: δ: 6.82 (d, J = 8.0, 2H), 6.78 (dd, J = 8.2, 2.3, 2H), 6.67 (dd, J = 8.1, 1.9, 1H), 6.63 (d, J = 1.8, 1H), 6.60 (dd, J = 8.0, 1.8, 2H), 6.59 (d, J = 1.9, 1H), 6.54 (d, J = 1.8, 2H), 2.75 (dd, J = 13.5, 5.0, 2H), 2.54 (dd, J = 13.5, 7.1, 2H), 2.40 (dd, J = 13.6, 7.6, 2H), 2.30 (dd, J = 13.5, 9.2, 1H), 1.76 (dd, J = 13.3, 6.7, 2H), 1.75 (dd, J = 13.0, 6.6, 2H), 0.85 (dd, J = 6.6, 2.5, 6H).

<sup>1</sup>*H* NMR (400 MHz, CDCl<sub>3</sub>) spectral data of **TL-3**:  $\delta$ : 7.46 (2H, d, J = 7.3, H-2''/2''' and 6''/6'''), 7.38 (2H, t, J = 7.4, H-3''/3''' and H-5''/5'''), 7.32 (1H, t, J = 7.2, H-4''/4'''), 6.81 (2H, d, J = 9.3, Ar-H), 6.79 (2H, d, J = 8.3, Ar-H), 6.69 (1H, d, J = 1.6, Ar-H), 6.64 (1H, dd, J = 9.4, 1.5, Ar-H), 6.57 (1H, dd, J = 8.1, 1.6, Ar-H), 5.14 (4H, s, H-7''/7)', 2.75 (2H, dd, J = 13.4, 4.9, H-7/7'), 2.57 (2H, dd, J = 13.6, 6.7, H-7/7'), 2.40 (2H, dd, J = 13.6, 7.8, H-7/7'), 2.30 (2H, dd, J = 13.4, 9.3, H-7/7'), 0.86 (3H, d, J = 7.2, H-9/9'), 0.84 (3H, d, J = 6.8, H-9/9') (see supporting information, Figure 15).



**Figure 13.** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) Spectrum of Compound 2 modified (**TL-2**) (schineolignin B modified)

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.76– 6.57 (5H, *m*, ArH), 3.86–3.81 (12H, *s*, 3 x OCH<sub>3</sub>), 2.56 (2H, *dd*, *J* = 13.5, 6.7, H-7/7<sup>7</sup>), 2.40 (2H, *dd*, *J* = 13.5, 7.8, H-7/7<sup>7</sup>), 2.30 (3H, *s*, CH<sub>3</sub>-CO), 1.79 – 1.73 (2H, *m*, H-8/8<sup>7</sup>), 0.83 (6H, *d*, *J* = 6.6, H-9/9<sup>7</sup>).



**Figure 14.** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) Spectrum of Compound **3** (mixture of *meso*-dihydroguaiaretic acid and *threo*-dihydroguaiaretic acid)

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.82 (2H, *d*, *J* = 8.0), 6.78 (2H, *dd*, *J* = 8.2, 2.3), 6.67 (1H, *dd*, *J* = 8.1, 1.9), 6.63 (1H, *d*, *J* = 1.8), 6.60 (2H, *dd*, *J* = 8.0, 1.8), 6.59 (1H, *d*, *J* = 1.9), 6.54 (2H, *d*, *J* = 1.8), 5.44 (1H, OH), 2.75 (2H, *dd*, *J* = 13.5, 5.0), 2.54 (2H, *dd*, *J* = 13.5, 7.1), 2.40 (2H, *dd*, *J* = 13.6, 7.6), 2.30 (2H, *dd*, *J* = 13.5, 9.2), 1.76 (2H, *dd*, *J* = 13.3, 6.7), 1.75 (2H, *dd*, *J* = 13.0, 6.6), 0.85 (6H, *dd*, *J* = 6.6, 2.5).



Figure 15. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) Spectrum of Compound **3** modified (TL-**3**) (mixture of *meso*-dihydroguaiaretic acid and *threo*-dihydroguai aretic acid modified)

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.46 (2H, *d*, *J* = 7.3, H-2<sup>''</sup>/2<sup>'''</sup> and 6<sup>''</sup>/6<sup>'''</sup>), 7.38 (2H, *t*, *J* = 7.4, H-3<sup>''</sup>/3<sup>'''</sup> and H-5<sup>''</sup>/5<sup>'''</sup>), 7.32 (1H, *t*, *J* = 7.2, H-4<sup>''</sup>/4<sup>'''</sup>), 6.81 (2H, *d*, *J* = 9.3, Ar-H), 6.79 (2H, *d*, *J* = 8.3, Ar-H), 6.69 (1H, *d*, *J* = 1.6, Ar-H), 6.64 (1H, *dd*, *J* = 9.4, 1.5, Ar-H), 6.57 (1H, *dd*, *J* = 8.1, 1.6, Ar-H), 5.14 (4H, *s*, H-7<sup>''</sup>/7<sup>'''</sup>), 2.75 (2H, *dd*, *J* = 13.4, 4.9, H-7<sup>'</sup>/7<sup>''</sup>), 2.57 (2H, *dd*, *J* = 13.6, 6.7, H-7<sup>'</sup>/7<sup>''</sup>), 2.40 (2H, *dd*, *J* = 13.6, 7.8, H-7<sup>'</sup>/7<sup>''</sup>), 2.30 (2H, *dd*, *J* = 13.4, 9.3, H-7<sup>'</sup>/7<sup>''</sup>), 0.86 (3H, *d*, *J* = 7.2, H-9<sup>'</sup>/9<sup>'</sup>), 0.84 (3H, *d*, *J* = 6.8, H-9<sup>'</sup>/9<sup>'</sup>).

#### **RESULTS AND DISCUSSION**

Three structural transformation process are presented in this article; and corresponds to the first report of this type of structural modification of lignans isolated from *Nectandra* species. A direct method was developed for the conversion of compound 1 to furan-type lignan. Additionally, the structural transformation of compounds 2 [14] (benzylation); and compound 3 [15] (acetylation). The spectroscopic data comparison between the initial

and the transformed compound showed formation of derivatives compounds (*see supporting information* for details). Interestingly, few reports describe dehydrogenation, benzylation or acetylation of natural products isolates; to our knowledge the direct structural transformation of lignan compounds isolated from *Nectandra* species has yet to be documented.

Comparison of spectroscopic data between the starting material (veraguensin and galgravin) and the product (**TL-1**) show the absence some characteristics signals [such as: 4.42 (1H, d, J = 9.3, H-7), 5.14 (1H, d, J = 8.6, H-7'), and 4.52 (2H, d, J = 6.4, H-7/7')], allow suggest the formation of **TL-1**.

The compound **TL-2** has a signal 2.30 (3H, s, CH<sub>3</sub>-CO), among others; which it is characteristic of the formation of the product.

The compound **TL-3** has a signal 5.14 (4H, *s*, H-7"/7"), among others; which it is characteristic of the formation of the product. Additionally, the compound formed is absent the signal generated by the hydroxyl group [5.44 (1H, *s*, OH)].

#### REFERENCES

- J. M. Barbosa-Filho, M. Yoshida, O. R. Gottlieb, Phytochemistry. 28, 1991, (1989).
- L. Chérigo, V. Polanco, E. Ortega-Barria, M. V. Heller, T. L. Capson, L. C. Rios, Nat. Prod. Res. 19, 373, (2005).
- 3. A. A. da Silva Filho, S. Albuquerque, M. L. e. Silva, M. N. Eberlin, D. M.

Tomazela, J. K. Bastos, J. Nat. Prod., 67, 42, (2004).

- S. R. Farias-Moreno, A. Arnobio, J. José de Carvalho, A. L. Nascimento, M. O. Timoteo, B. Olej, E. K. Rocha, M. Pereira, M. Bernardo-Filho, L. Querino de Araújo Caldas, Biol. Res. 40, 131, (2007).
- J. C. Moro, J. B. Fernandes, P. C. Vieira, M. Yoshida, O. R. Gottlieb, H. E. Gottlieb, Phytochemistry, 26, 269, (1987).
- B. Agius, M. Setzer, S. Stokes, T. Walker, W. Haber, W. Setzer, Int. J. Essen. Oil Ther. 1, 167, (2007).
- J. G. Rohwer. Lauraceae: Nectandra. Flora Neotropica, Monograph 60, in Flora Neotropica Monograph. vol. 60, T. N. Y. B. Garden, Ed., ed New York, pp. 1-332, 1993.
- 8. J. Chen, W. Li, H. Yao, J. Xu, Fitoterapia. 103, 231, (2015).
- 9. M. Saleem, H. J. Kim, M. S. Ali, Y. S. Lee, Nat. Prod. Rep, 22, 696, (2005).
- L. Dalla Via, E. Uriarte, E. Quezada, A. Dolmella, M. G. Ferlin, O. Gia, J. Med. Chem. 46, 3800, (2003).
- 11. R. Nakamura, Y. Obora, Y. Ishii, Tetrahedron. 65, 3577, (2009).
- 12. H. S. P. Rao, S. Senthilkumar, J. Chem. Sci. 113, 191, (2001).
- 13. L. McMaster, W. Bruner, Ind. Eng. Chem. 28, 505, (1936).
- 14. M. Miyazawa, H. Kasahara, H. Kameoka, Phytochemistry. 46, 1173, (1997)
- Y. B. Xue, Y. L. Zhang, J. H. Yang, X. Du, J. X. Pu, W. Zhao, X. N. Li, W. L. Xiao, H. D. Sun, Chem. Pharm. Bull. 58, 1606, (2010)-