STRUCTURE AND THERMAL BEHAVIOR OF TANNINS FROM Acacia dealbata BARK AND THEIR REACTIVITY TOWARD FORMALDEHYDE

JUSTO LISPERGUER^{1,2*}, YANINA SARAVIA², EMILIO VERGARA³

¹Facultad de Ciencias, Departamento de Química, Universidad del Bio-Bio, Av Collao 1202 Concepción, Chile ²Centro de Investigación de Polímeros Avanzados, CIPA. Conicyt-Regional R08C1002. Av. Collao 12, Concepción, Chile. ³Centro de Investigación en Tecnologías de la Construcción, Universidad del Bio-Bio. Av. Collao 1202, Concepción, Chile.

ABSTRACT

The objective of this study was to evaluate the tannin extraction potential taken from the bark of *Acacia dealbata*. This tannins were extracted with water at 90°C. An extraction yield of 17.2% solids with a Stiasny number of 82% was obtained from the bark extract of *Acacia dealbata*. The structure was studied by Fourier transform infrared spectroscopy (FTIR). The thermal behavior of tannins was studied by DSC and a glass- transition temperature (T_g) of 116,77°C was determined. The thermal stability of tannins was studied by TGA. At 196.91°C the decomposition is 3.7% and the maximum of the weight loss rate (DTG) of the degradation was 257.77°C. The curing with formaldehyde showed an exotherm reaction in the range of 100-120°C, which is similar to tannins of other species.

Keywords: Tannins, Acacia dealbata bark, Structure of tannins, Thermal behavior.

INTRODUCTION

The potential of tannins, particularly those extracted from bark, has long been recognized to substitute more expensive petrochemical-derived components such as phenol-formaldehyde, resorcinol-formaldehyde or ureaformaldehyde, in adhesives for wood gluing applications¹⁻⁶. Tannins are natural polyphenolic materials, composed mostly of flavan-3ol repeating units and smaller fractions of polysaccharides and sugars. These polyphenolic materials can be hardened by reaction with formaldehyde or hexamethylenetetramine (HEXA), as crosslinking agents^{7,8}. Examples of tannin use in thermoset adhesive applications include partial or full replacement of phenol formaldehyde resin in hot –pressing of products such as plywood and composite panels, and the partial substitution of resorcinol in adhesive resins^{9,10}.

Studies that report the behavior of these adhesives for the manufacture of molding powers or fiber composite materials are scarce in the open literature¹¹.



Figure 1. Flavan- 3ol repeating unit in polyflavonoid tannins.

The reaction of HEXA with tannins was studied by Pizzi, who showed that HEXA is not a formaldehyde-yielding hardener and thus it lead to cured products with low formaldehyde emissions. The crosslinking reactions proceed by formation of reactive HEXA fragments or intermediates that react with phenolic nuclei of the polyflavonoid tannins¹.

The most widely used as industrial tannins are obtained from the wood and bark of the quebracho tree (*Schinopsis balansae*, Argentine) and from the bark of mimosa (*Acacia mearnsii*, Brazil and South Africa). Resorcinolictype tannins such as those derived of wattle or quebracho species tend to only find applications in hot pressing¹¹. Other tannins obtained from pine species with phloroglucinolic structures, have and additional hydroxyl group on the A- ring of the tannin flavonoid unit giving greater reactivity allowing use in ambient temperature curing (cold set adhesives). Phloroglucinolic-type tannins, compared with resorcinolic type tannins, have found little commercial use, due to undesirable characteristic as high reactivity with formaldehyde and high viscosity¹ Phenol-formaldehyde based wood adhesives still today dominant in European markets. However, the necessity to diminish petrochemicals consumption stimulate the search of alternative environmentally safe adhesives. The antimicrobial properties of tannins make their application in wood composites and adhesives industry very attractive¹².

In this work, we have investigated about the isolation, structure and thermal behavior of tannins from *Acacia dealbata* bark. We also studied the reactivity of the tannins toward formaldehyde.

EXPERIMENTAL

Extraction method

100 g of ground bark from *Acacia dealbata* and 1700 mL of water were placed in a flask and heated at 90° C with mechanical stirring during 6 hours. Once extraction time was completed, the extraction liquor was separated from the solid residue by filtration. The solid residue was then washed four times and washing waters were evaporated in an oven at 100° C overnight.

Determination of polyphenols by the Stiasny reaction

The Stiasny number reaction was used to determine the polyphenol content of extracts. Fifty milliliters of (15% w/w) tannin solution was pipeted into a 250 ml flask. 10 ml of aqueous formaldehyde (37% w/w) and 5 ml of hydrochloric acid solution (10M) were added and the mixture was heated under reflux for 30 min. The reaction mixture was filtered through a sintered glass filter (porosity 2) whilst it was still hot. The precipitate was then dried in an oven at 105° C to constant weight. The Stiasny number is the ratio of the oven-dried weight of the precipitate to the total dissolved solid content of the tannin extract expressed as a percentage.

FT-IR analysis

Tannins from dried extract of *Acacia dealbata* were analyzed by Fourier transform infrared spectroscopy (FT-IR) on a Perkin Elmer FT-112 Spectrum Two instrument to characterize functional groups of tannins.

TGA and DSC analysis

To determine the thermal behavior of tannins, a dynamic test was carried out with a TGA Q 50. The samples, with an average mass of 8 mg, were heated from 25 a 600° C at a heating rate of 10° C/min under a nitrogen atmosphere. The thermal stability of tannins was studied.

The glass-transition temperature (T_g), was studied with a DSC 822e Mettler Toledo equipped with a data analyzed software STARe. For this purpose, the ASTM 3418 was applied within a range of 25 and 200° C at a heating rate of 20° C/min, under a nitrogen atmosphere.

Curing reaction of tannins adhesives

Tannin solution was prepared and 10% of powdered paraformaldehyde, by weight of dry tannin extract, was added. NaOH was added until a pH of 11. The sample was sealed under air in 120 μ L medium pressure stainless steel crucible with a Viton O-ring which can withstand pressures up to 2 MPa. The temperature of the DSC was scanned from 25° C to 250°C at a heating rate of 10°/min under a nitrogen atmosphere.

RESULTS AND DISCUSSION

Solids and polyphenols content of the bark extract

An extraction yield of 17.2% solids was obtained from the bark extract of *Acacia dealbata*. This value is slightly less than those mentioned for mimosa and *Pinus radiata* barks (i.e. 42.5% and 20%, respectively)^{13,14}

The mean values of polyphenols content determined by the Stiasny method, was 82%. This value is also similar to that found for other species¹⁵.

Grace pomace also been used to obtain tannins. These extract have a considerable proportion of nonphenolic materials, mainly simple sugars and polymeric carbohydrates with low Stiasny numbers. The yield extraction in water at 120°C was 22.3% (oven dry mass) and the Stiasny number ranged between 32 to 55^{16,17}.

Other researchers have reported the tannins extraction yield of Douglas fir bark of 14% in case of urea/sulphite solution.¹⁸

FT-IR analysis of tannins

In Fig. 2 the FT-IR spectrum of the tannins from *Acacia dealbata* bark extract are shown. The area between 500 and 4000 cm⁻¹ was analyzed.



Figure 2. FT-IR spectra of tannins from Acacia dealbata bark.

In Fig. 2, it can be clearly observed a wide band at 3306 cm⁻¹ which shows the presence of the phenolic hydroxyl groups (OH stretching vibrating). The 2919 and 2850 cm⁻¹ absorption bands were attributed to C-H and CH₂ vibrations of aliphatic hydrocarbon. The 1608.3, 1512 and 1449 cm⁻¹ absorption bands, were attributed to aromatic ring stretching vibration. The 1034.8 cm⁻¹ absorption band, was attributed to C-O stretching vibration. Tannins from Taiwan acacia bark shows only one band at 2933 cm⁻¹ attributed to C-H and CH₂ vibration.¹⁹ Tannins of other species such as *Pachira quinata* and *Pinus caribea* show very similar FT-IR spectra to *Acacia dealbata*.²⁰

Thermal characterization

DSC analysis

DSC is the most widely accepted method for determining the glass transition temperature of natural and synthetic polymers and generally to study the thermal behavior of a polymer. Figure 3 shows the DSC curve for tannin from *Acacia dealbata* bark extract obtained at a heating rate of 10°/min.

The T_g of dry tannin is often more difficult to detect than in a synthetic polymer, due the complex structure of tannin and sometimes only is possible to detect the range of the change in the curve.

The T_g of the tannin was determined from the change in the heating curve and it is located at 116,77° C, the same region reported for tannins of other species as quebracho tannins where the T_g was located at 126°C.⁴ This value is also similar to the T_g of lignin of *Eucaliptus* and *Pinus radiata*²¹.

TGA analysis measurement

In order to study the thermal stability of the tannins, runs by means of TGA were conducted. .



Figure 3. DSC of Acacia dealbata tannin.



Figure 4. Decomposition curves of Acacia dealbata tannin.

As shown in Figure 4, the decomposition profile of the tannin occurred in one step. At 196.91°C, the decomposition is only a 3.7%, which can be attributed to the loss of moisture and gases absorbed and does not involve degradation of the tannins. The maximum of the weight loss rate (DTG) of the degradation, was 257.77°C.

The complex condensed aromatic structure of tannin leads to high thermal resistance. Although the degradation is almost complete at 600°C, tannin shows a remaining weight of about 44%.

Curing reaction of tannin-formaldehyde adhesives

Figure 5 shows the DSC scan obtained at a heating rate of 10°C for the curing reaction of tannin with paraformaldehyde. The characteristic curing reaction shows only one peak of the exothermic reaction in the range of 100-120°C with a peak at 110,74°C.

The curing reaction of tannin occurs at lower temperature than phenol formaldehyde resins (145-150°C) and is more similar to the reaction curing of phenol- resorcinol formaldehyde (60-120°C).²²

The endotherm in the range of 200-220°C shows the thermal degradation of tannin, also shown in the figure 4.

CONCLUSIONS

In this study, the structure, thermal properties and curing reactions of *Acacia dealbata* bark tannins were analyzed. An extraction yield of 17.2% solids was obtained from *Acacia dealbata* bark. This is slightly less than those mentioned for *mimosa* and *Pinus radiata* barks. The polyphenols content determined by the Stiasny number was 82% and it is similar to that found for other species. TGA studies show a high thermal resistance of tannins. The decomposition starts at 196.91°C with a DTG at 257.77°C. DSC studies show a T_g of tannin of 116.77°C similar to other lignocellulosic materials as lignin. The curing reactions of tannin with formaldehyde occurs in the range of 100-120°C with a peak at 110.74°C, which is similar to tannins of *Pinus radiata* and

mimosa. The study of the properties of tannins of *Acacia dealbata* shows that it is possible to use these tannins in the formulation of adhesive resins reinforced with phenol formaldehyde.



Figure 5. DSC thermogram of the curing reaction of tannin-formaldehyde adhesive.

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REFERENCES

- 1. A.Pizzi, J. Adhesion Sci. Technol. 20, 829, (2006).
- 2. M.H.Alma, S.S.Kelley, *Polym.Degrad.Stab.* 68, 413, (2003).
- 3. E.Roffael, B.Dix, J.Okum, Holz als Roh und Werkstoff 58, 301, (2000).
- M.Mosiewicki, M.I.Aranguren, J.Borrajo, J.Appl. Polym.Sci. 91, 3074, (2004).
- 5. M.Fechtal, B.Riedl, *Holzforschung* 47, 349, (1993).
- 6. Y. Lu, Q.Shi, Holz als Roh und Werkstoff 53, 17, (1995).
- A.Pizzi in Handbook of Adhesive Technology, A.Pizzi, K.L.Mittal eds. Marcel Dekker, New York, 2003.
- 8. F.Pichelin, C.Kamoun, A.Pizzi, Holz als Roh und Werkstoff 57, 305, (1999).
- P.M.Stefani, C.Peña, R.A.Ruseckaite, J.C.Piter, I Mondragon, *Bioresource Technology* 99, 5977, (2008).
- 10. W.J.Lee, W.C.Lan, Bioresource Technology 97, 257, (2006).
- R. Nico, Plásticos de tanino de quebracho, Laboratorio de Ensayo de Materiales e Investigaciones Tecnológicas-LEMIT, Report 38, Series II, La Plata, Argentina, (1951).
- A.Pizzi in Wood Adhesives: Chemistry and Technology, A.Pizzi ed.Marcel Dekker, New York, 1983.
- 13. A. Scalbert, Phytochem. 30, 3875, (1991).
- W.J.Grigsby, S.J.Hill, C.D.McIntosh, J.Wood Chem. Technol. 23, 179, (2003).
- 15. S.Ohara, Y.Yasuta, H.Ohi, Holzforschung 57, 145, (2003).
- 16. L.Ping, A.Pizzi, Z. D. Guo, N.Brosse, Ind. Crops Prod. 40, 13, (2012)
- 17. L.Ping, A.Pizzi, Z.D. Gu0, N.Brosse, Ind. Crops Prod. 34, 907, (2011)
- F.Bertaud, S.Tapin-Lingua, A.Pizzi, P.Navarrete, M.Petit-Conil, Cellulose Chem. Technol. 46, (7-8), (2012)
- 19. W.J.Lee, W.C. Lan, Bioresource Technol. 97, 257, (2006)
- J. Aguilar, J. Jaen, A.Vargas, P.Jiménez, L. Vega, J.Herrera, H.Borbón, R, Soto, *Tecnología en Marcha* 25, (4), 15 (2012)
- 21. J.Lisperguer, P.Pérez, S Urizar, J.Chil.Chem.Soc. 54, 460, (2009)
- 22. J. Lisperguer, C. Droguett, B. Ruf, J. Chil. Chem. Soc. 50, 375, (2005).